SIX GUAIANOLIDES FROM STYLOTRICHIUM ROTUNDIFOLIUM*

FERDINAND BOHLMANN,† ANTOINETTE SUWITA,† HAROLD ROBINSON; and ROBERT M. KING;

†Institute for Organic Chemistry, Technical University of Berlin, D-1000 Berlin 12, West Germany;‡ Smithsonian Institution, Washington, DC 20560, U.S.A.

(Received 3 November 1980)

Key Word Index—Stylotrichium rotundifolium; Compositae; Eupatorieae; sesquiterpene lactones; guaianolides; germacranolides; geranylnerol derivative.

Abstract—The investigation of Stylotrichium rotundifolium afforded the epoxide of β -sesquiphellandrene, a new derivative of geranylnerol and six guaianolides not isolated previously. Furthermore, two germacranolides also present in a Lasiolaena species and several known compounds were isolated. The structures were elucidated by detailed ¹H NMR investigations. The chemotaxonomic situation is discussed briefly.

INTRODUCTION

The small Brazilian genus Stylotrichium is placed in the diverse Gyptis group (Compositae, tribe Eupatorieae) [1]. In continuation of our chemotaxonomic studies of the Eupatorieae, we have now investigated S. rotundifolium Mattf. to see whether the chemistry gives any indication of relationship to other genera placed in the Gyptis group. The isolation of several sesquiterpene lactones and of a new diterpene shows there is a close relationship to Lasiolaena, which also belongs to the Gyptis group [1].

RESULTS AND DISCUSSION

The roots of S. rotundifolium afforded sesquiphellandrene, dammaradienyl acetate, the phydroxyacetophenone derivatives 1, 2 [2], 3 [2] and the dehydronerolidol derivative 10 [3], while the aerial parts gave germacrene D, bicyclogermacrene, α-humulene, caryophyllene, β -sesquiphellandrene, the angelate 4 [4], taraxasterol and its acetate, the germacranolides 11 and 12 [5] as well as several new compounds, namely the epoxide of β -sesquiphellandrene (5), the geranylnerol derivative 6 and the guaianolides 13-18. The structure of 5 was deduced from the ¹H NMR data, which were very close to those of β -sesquiphellandrene (see Experimental). However, the signals of the isopropylidene group were replaced by two singlets at 1.32 and 1.27 ppm and a triplet at 2.71 (J = 6 Hz). The latter was obviously the signal of an epoxide proton. The relative stereochemistry at C-10 could not be assigned. The structure of 6 also followed from the ¹H NMR data (Table 1), those of the products obtained by manganese dioxide oxidation (7 and 8) and of the corresponding diacetate 9. As could easily be deduced from these data, a geranylnerol derivative must be present with an additional hydroxyl group at C-18 or C-19 and a keto group at C-13 since the methyl groups at C-15 and

the proton at C-14 were shifted downfield. From the data of the aldehydes 7 and 8 the position of the second oxygen function could be deduced. As the H-12 signal was not shifted downfield in the spectrum of 8, the aldehyde group must be placed at C-7. The stereochemistry of the 2,3double bond followed from the chemical shifts of H-20 in the spectrum of 7 and 8. 6 is closely related to a corresponding diterpene isolated from a Lasiolaena species [5]. The ¹H NMR data of 13–18 (Table 2) showed that all six lactones were closely related to each other. 16 obviously was the 5'-O-acetate of agriantholide (19), isolated from an Agrianthus species [6]. Very similar compounds were isolated from Liatris and Eupatorium species [7, 8]. The ¹H NMR data therefore were also similar. In the spectra of 13-15 the presence of a 10,14double bond clearly followed from the typical broadened singlets between 5.4 and 5.25 ppm, while the presence of the corresponding epoxide in 16-18 was indicated by the two doublets around 3.0 ppm. Furthermore, an olefinic signal about 5.7 ppm and a broadened singlet at 1.0–2.0 ppm are typical for guaianolides with a 3,4-double bond. Spin decoupling showed that in all cases 6,12lactones were present. Starting with the H-7 signals the protons for H-1 through H-3, H-5 through H-9 and H-13 could readily be assigned. Surprisingly there were, however, two groups with different couplings $J_{8,9}$. Those lactones with a 9-hydroxy function showed a coupling of about 5 Hz and those with a 9-ester group a 2 Hz coupling. Furthermore, in the spectrum of 14 and 15 a downfield shift of H-1 was observed. The small couplings for $J_{7,8}$ in the spectra of all lactones indicated a β -orientation for the oxygen functions at C-8 in all cases. The stereochemistry at the other centres was obviously the same too in all compounds and corresponded with that of the lactones with known stereochemistry [6-8]. The nature of the oxygen function also clearly followed from the ¹H NMR data. Most probably, 16-18 are 10,14α-epoxides as the H-1 signals of 15-18 were at lower field like the H-1 signal in agriantholide.

13-15 were derived from precupatundin [8] and 16-18 from agriantholide ([6]; see Note Added in Proof). The chemistry of *Stylotrichium* is very close to that of

^{*}Part 345 in the series "Naturally Occurring Terpene Derivatives". For Part 344 see Bohlmann, F. and Fritz, G. (1981) Tetrahedron Letters 95.

Table 1. ¹H NMR spectral data of compounds 6-9 (270 MHz, CDCl₃, TMS as internal standard)

	6	7	8	9
H-1	4.08 br d	9.91 d	9.92 d	4.55 br d
H-2	5.48 br t	5.91 br d	5.96 br d	5.40 br t
H-4 H-5	2.20 m	$ \begin{array}{c} 2.64 \ t \\ 2.36 \ dt \end{array} $	2.80 m	2.20 m
H-6	5.33 br t	5.33 br t	6.42 br t	5.37 br t
H-8 H-9	2.20 m	2.20 m	2.15 m } 2.24 br dt }	2.20 m
H-10	5.26 br t	5.24 br t	5.19 br t	5.23 br t
H-12 H-14 H-16	3.05 br s 6.11 qq 1.89 d	3.06 br s 6.11 qq 1.89 d	3.04 <i>br s</i> 6.10 <i>qq</i> 1.89 <i>d</i>	3.04 br s 6.10 qq 1.88 d
H-17 H-18	2.15 d 1.63 br s	2.14 <i>d</i> 1.62 <i>br s</i>	2.14 d 1.60 br s	2.14 d 1.63 br s
H-19	$4.10 \ br \ s$	4.13 br s	10.09 s	4.59 s
H-20	1.77 br s	1.99 d	2.01 d	1.76 br s
OAc	_	_	_	2.06 s 2.04 s

J (Hz): 1,2 = 5, 6 = 9, 10 = 7; 14, 16 = 14, 17 = 1.3.

Lasiolaena [5] and Agrianthus [6]. The constituents of the other genera placed in the Gyptis group so far investigated, however, are of different types. Only Trichogonia [10] and Conocliniopsis [11] also contain sesquiterpene lactones, mainly furanoheliangolides.

EXPERIMENTAL

The air-dried plant material (voucher RMK 8149) was extracted with Et₂O-petrol (1:3) and the resulting extracts were separated first by column chromatography (Si gel)

and further by repeated TLC (Si gel). Known compounds were identified by comparing the IR and 1H NMR spectra with those of authentic material. The roots (80 g) afforded 15 mg β -sesquiphellandrene, 10 mg dammaradienyl acetate, 3 mg 1, 15 mg 2, 10 mg 3 and 30 mg 10, while the aerial parts gave 10 mg germacrene D, 40 mg β -sesquiphellandrene, 20 mg bicyclogermacrene, 20 mg α -humulene, 10 mg taraxasterol and 50 mg of its acetate, 20 mg caryophyllene, 50 mg 4, 5 mg 5 (Et₂O-petrol, 1:20), 11 mg 6 (Et₂O-petrol, 3:1), 10 mg 11, 3 mg 12, 4 mg 13 and 14 (CHCl₃-Me₂CO, 4:1), 3 mg 15 (CHCl₃-Me₂CO, 3:2), 10 mg

Table 2. ¹H NMR spectral data of compounds 13-18 (270 MHz, CDCl₃, TMS as internal standard)

	13	14	15	16	17	18
H-1	3.23 br dd	3.63 br dd	3.69 br dd	2.67 dd	2.85 dd	2.65 dd
H-2	4.98 br d	4.79 br d	4.77 br d	4.78 br d	4.57 br dd	4.83 br d
H-3	5.72 dq	5.65 dq	5.60 dq	5.65 dq	5.84 dq	5.63 br s
H-5	2.88 br dd	2.85 br dd	2.84 br dd	2.81 br dd	2.76 br dd	2.83 br da
H-6	4.61 dd	4.98 dd	5.08 dd	5.15 dd	4.94 dd	5.16 dd
H-7	2.98 dddd	3.29 dddd	3.29 dddd	3.24 dddd	3.17 dddd	3.28 dddd
H-8	4.49 dd	5.88 dd	5.58 dd	5.50 dd	6.00 dd	5.54 dd
H-9	5.19 br d	4.72 br d	4.69 br d	3.96 d	5.32 d	3.84 d
H-13	6.41 d	6.29 d	6.28 d	6.32 d	6.24 d	6.32 d
H-13'	5.46 d	5.51 d	5.57 d	5.56 d	5.54 d	5.55 d
H-14	5.41 br s	5.38 br s	5.33 br s	3.01 d	3.00 d	2.95 d
H-14'	5.36 br s	5.30 br s	5.26 br s	2.86 d	2.88 d	2.90 d
H-15	1.90 br s	1.94 br s	1.95 br s	1.99 br s	2.03 br s	1.99 br s
OCOR	7.29 q	7.15 q	6.96 q	7.14 q	7.20 q	6.96 q
	2.03 d	1.97 d	1.89 d	1.94 d	1.95 d	1.90 d
	400 1 -	4.85 d	4.38 d	4.87 d (4.90 d {	4.34 d
	4.99 br s	4.78 d {	4.27 d {	4.78 d {	4.80 d	4.27 d
OAc	2.09 s	2.03 s	_	2.01 s	2.10 s	
					1.99 s	

J (Hz): 1,2 = 7; 1,5 = 9.5; 2,3 \sim 2; 3,15 = 1; 5,6 = 10.5; 6,7 = 9; 7,8 \sim 3.5; 7,13 = 3.5; 7,13' = 3; 8,9 = 5 (14–16, 18; 13 and 17: 1.8); 16–18: 14,14' = 4.5; 3',4' = 7; $5_1'$, $5_2'$ = 12.5.

16 and 3 mg 17 (CHCl₃-Me₂CO, 6:1) and 5 mg 18 (CHCl₃-Me₂CO, 3:2).

Epoxy-β-sesquiphellandrene (5). Colourless oil; IR $v_{\rm cm}^{\rm cmla}$ cm⁻¹: 2940, 2860, 1645, 1470, 1385, 1365, 1225, 890; MS m/z (rel. int.): 220.183 (M⁺, 13) (C₁₅H₂₄O), 202 (M - H₂O, 10), 187 (202

 $(CDCl_3)$: 0.88 (d, H-14, J = 7 Hz), 1.27 and 1.32 (s, H-12, 13), 2.71 (t, H-10, J = 6 Hz), 4.76 $(br \ s, H-15)$, 6.17 (dd, H-1, J = 10, 2.5 Hz), 5.68 $(br \ d, H-2, J = 10 Hz)$.

19-Hydroxy-13-oxo-geranylnerol (6). Colourless gum: IR $v_{\text{max}}^{\text{CCl}_4}$ cm⁻¹: 3620 (OH), 1690, 1625 (C=CCO); MS m/z (rel. int.) (CI, *i*-butane): 321 (M + 1, 4), 303 (321 - H₂O, 44), 285 (303 - H₂O, 44), 83 (MeC=CHCO⁺, 100). 10 mg 6 in 2 ml Et₂O was stirred for 1 hr with 50 mg MnO₂. TLC (Et₂O) afforded 6 mg 7 and 3 mg 8. 7: Colourless gum, IR $v_{\text{max}}^{\text{CCl}_4}$ cm⁻¹: 3630 (OH), 2760, 1685 (CHO), 1685, 1625 (C=CCO); MS m/z (rel. int): 318.220 (M⁺, 0.3), 83 (Me₂C=CHCO⁺, 100); 8: Colourless gum, IR $v_{\text{max}}^{\text{CCl}_4}$ cm⁻¹: 1680, 1620 (C=CCO); MS m/z (rel. int.): 316.204 (M⁺, 0.2), 83 (MeC=CHCO⁺, 100). 1 mg 6 was heated for 30 min in 0.1 ml Ac₂O at 70°. TLC (Et₂O-petrol, 1:1) afforded 1 mg 9, colourless gum (¹H NMR data, see Table 1).

8 β -Hydroxy-9 β -[5-acetoxytiglinoyloxy]-preeupatundin (13). Colourless gum; IR $\nu_{\text{max}}^{\text{CHZ}_3}$ cm $^{-1}$: 3540 (OH), 1780 (γ -lactone), 1740 (CO₂R); MS m/z (rel. int.) (CI, i-butane): 419 (M + 1, 8), 391 (419 - CO, 11), 279 (419 - O=C=CR, 8), 117 (HO₂CC(CH₂OH)=CHMe + 1, 100), 99 (117 - H₂O, 71).

$$[\alpha]_{24^{\circ}}^{\lambda} = \frac{589}{+14} \frac{578}{+16} \frac{546}{+18} \frac{436 \text{ nm}}{+34} (c = 0.3, \text{ CHCl}_3).$$

9 β -Hydroxy-8 β -[5-acetoxytiglinoyloxy]-preeupatundin (14). Colourless gum; IR $\nu_{max}^{\text{CHCl}_3}$ cm⁻¹: 3620 (OH), 1765 (γ -lactone), 1730 (CO₂R); MS m/z (rel. int.): 418 (M⁺, 0.2), 400, 152 (M - H₂O, 2) (C₂₂H₂₄O₇), 242 (400 - RCO₂H, 48), 141 (RCO⁺, 94), 81 (141 - HOAc, 100); [α]_{436 nm} + 9° (c = 0.4, CHCl₃).

9 β -Hydroxy-8 β -[5-hydroxytiglinolyoxy]-preeupatundin (15). Colourless gum; IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm $^{-1}$: 3600 (OH), 1770 (γ -lactone), 1730 (CO₂R); MS m/z (rel. int.): 376 (M $^+$, 0.3), 358.142 (M $^-$ H₂O, 1.5) (C₂₀H₂₂O₆), 260 (M $^-$ RCO₂H, 7), 242 (260 $^-$ H₂O, 20), 99 (RCO $^+$, 100). [α]_D +4 $^\circ$ (c = 0.1, CHCl₃).

5'-Acetoxy-agriantholide (16). Colourless gum; IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm⁻¹: 3600 (OH), 1770 (γ -lactone), 1720 (CO₂R); MS (CI, *i*-butane): 435 (M + 1, 4), 417 (435 - H₂O, 100), 399 (417 - H₂O, 66), 259 (417 - RCO₂H, 64), 241 (259 - H₂O, 63).

5'-Acetoxy-agriantholide-9-O-acetate (17). Colourless gum; IR $\nu_{\text{max}}^{\text{CHC1}_3}$ cm $^{-1}$: 3600 (OH), 1760 (γ-lactone), 1730 (CO₂R); MS (CO, i-butane): 477 (M + 1, 0.2), 459 (477 - H₂O, 100), 417

(477 - HOAc, 24), 399 (459 - HOAc, 20), 159 $(\text{RCO}_2\text{H} + 1, 14)$, 141 $(159 - \text{H}_2\text{O}, 12)$.

$$[\alpha]_{24^{\circ}}^{\lambda} = \frac{589}{+44} \frac{578}{+48} \frac{546}{+51} \frac{436 \text{ nm}}{+91} (c = 0.3, \text{ CHCl}_3).$$

5'-Hydroxy-agriantholide (18). Colourless gum; IR $V_{\rm max}^{\rm CHCl_3}$ cm $^{-1}$: 3600 (OH), 1760 (γ-lactone), 1725 (CO₂R); MS m/z (rel. int.): 392.147 (M $^+$, 1) (C₂₀H₂₄O₈), 276 (M $^-$ RCO₂H, 5), 258 (276 $^-$ H₂O, 10), 99 (RCO $^+$, 100), 81 (99 $^-$ H₂O, 70).

$$[\alpha]_{24^{\circ}}^{\frac{1}{2}} = \frac{589}{+8} \frac{578}{+9} \frac{546}{+10} \frac{436 \text{ nm}}{+12} (c = 0.4, \text{ CHCl}_3).$$

Acknowledgements—We thank Drs. Scott A. Mori and P. Alvim, Herbario Centro de Pesquisas do Cacau at Itabanu, Bahia, Brazil, for their help in plant collection and the Deutsche Forschungsgemeinschaft for financial support.

REFERENCES

- Robinson, H. and King, R. M. (1977) The Biology and Chemistry of the Compositae (Heywood, V. H., Harborne, J. B. and Turner, B. L., eds.) p. 455, 463. Academic Press, London.
- Bohlmann, F., Zdero, C., King, R. M. and Robinson, H. (1980) Phytochemistry 19, 1141.
- 3. Bohlmann, F. and Zdero, C. (1971) Chem. Ber. 107, 964.
- Bohlmann, F. and Ziesche, J. (1979) Phytochemistry 18, 1489.
- Bohlmann, F., Jakupovic, J., King, R. M. and Robinson, H. (1981) Phytochemistry 20, 1613.
- Bohlmann, F., Zdero, C., Robinson, H. and King, R. M. (1980) Phytochemistry 19, 1873.
- 7. Herz, W. and Sharma, R. P. (1976) J. Org. Chem. 41, 1248.
- Bohlmann, F., Mahanta, P. K., Suwita, A., Suwita, Ant., Natu, A. A., Zdero, C., Dorner, W., Ehlers, D., and Grenz, M. (1977) Phytochemistry 16, 1973.
- Bohlmann, F., Jakupovic, J., Schuster, A., King, R. M. and Robinson, H. (1981) Phytochemistry 20 (in press).
- Bohlmann, F., Zdero, C., Pickardt, J., Robinson, H. and King, R. M. (1981) Phytochemistry 20, 1323.
- Bohlmann, F., Zdero, C., King, R. M. and Robinson, H. (1979) Phytochemistry 19, 1547.

NOTE ADDED IN PROOF

The stereochemistry of agriantholide has to be changed at C-9 following detailed ¹H NMR studies on the lactones from Lasiolaena morii [9].